

Biomass and element pools of understory vegetation in the catchments of Čertovo Lake and Plešné Lake in the Bohemian Forest

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Abstract: This paper presents data on species composition, biomass, and element pools (C, N, P, Ca, Mg, Na, K, Al, Fe, Mn) of the understory vegetation of spruce forests in the catchments of lakes Čertovo jezero (CT) and Plešné jezero (PL) in the Bohemian Forest (Šumava, Czech Republic). *Calamagrostis villosa* was the most abundant species in the CT catchment, while *Vaccinium myrtillus* was the most abundant species in the PL catchment. The catchments weighted mean (CWM) of above-ground biomass of the understory vegetation was 288 and 723 g m⁻² in the CT and PL catchments, respectively. The significant difference in the biomass between the catchments was caused by the much higher abundance of *V. myrtillus* in the PL catchment. The CWM of below-ground biomass of the fine roots was 491 and 483 g m⁻² in the CT and PL catchments, respectively. The respective CWM element pools of biomass in the CT and PL catchments were: C (33 and 51 mol m⁻²), N (0.8 and 1.0 mol m⁻²), P (24 and 34 mmol m⁻²), Ca (53 and 113 mmol m⁻²), Mg (24 and 41 mmol m⁻²), Na (3.7 and 6.5 mmol m⁻²), K (83 and 109 mmol m⁻²), Al (50 and 42 mmol m⁻²), Fe (13.3 and 7.3 mmol m⁻²), and Mn (4.2 and 8.8 mmol m⁻²).

Key words: Norway spruce forest, understory vegetation, *Calamagrostis villosa*, *Vaccinium myrtillus*.

Introduction

Forest research has always tended to focus on the trees, while much less attention has been paid to understory components such as dwarf shrubs, herbs, grass, ferns and mosses (NILSSON & WARDLE, 2005). However, the importance of understory vegetation in forest ecosystems is probably comparable to that of the trees. The ecological importance of understory vegetation can be viewed at the vegetation and soil process scales. Understory vegetation can strongly influence tree seedling establishment and growth (GEORGE & BAZZAZ, 1999) and thus the dynamics of the whole forest stand. Soil processes affect belowground processes, such as decomposition and build-up of soil nutrients (JONASSON & SHAVER, 1999). Understory vegetation can also serve as an important pool of nutrients in forest ecosystems. For example, the annual aboveground production of understory vegetation was estimated to be between 25 and 32% of total aboveground biomass for a Norway spruce stand in boreal Fennoscandia (FINER et al., 2003). In the same stands, the annual uptake of C and N by understory vegetation represented as much as 22 and

56% of the annual net uptake of C and N by trees, respectively (FINER et al., 2003). Similar values were reported for Scots pine dominated forests from boreal Fennoscandia (HELMISAARI, 1995). According to some recent studies, understory vegetation can also affect and modify nutrient flow in forest ecosystems (RODENKIRCHEN, 1995; HUBER et al., 2004). It has been shown that grass understory vegetation can serve as an important sink of atmospheric nitrogen (HOLUB, 1999). Sward grass vegetation can also decrease soil acidity and the loss of base cations (FIALA et al., 2005). This suggests that some species of understory vegetation can partly eliminate negative processes associated with soil acidification and positively affect the reduction of nutrient losses from the soil (FIALA et al., 2005). Contradictory results were found by ŠANTRŮČKOVÁ et al. (2006), showing that vegetation cover of *Calamagrostis villosa* possibly contributed to high N flux from the litter in the spruce forest of the Bohemian Forest. On the whole catchment scale, N flux from the litter was similar to N input by atmospheric deposition. Based on these results, there is an obvious need for detailed studies on the role of understory vegetation

within the process of element cycling on the catchment scale.

The aim of this paper is to evaluate differences in the character, biomass and element pools of the understory vegetation in the spruce forest of two Bohemian Forest catchments of lakes Plešné jezero (PL) and Čertovo jezero (CT). Catchments of both lakes have been exposed to high deposition of S and N compounds during the last six decades, but have been partly recovering from this acid stress since the late 1980s (MAJER et al., 2003). Long term ecosystem research focusing on the process of ecosystem acidification and biological recovery has been carried out in both catchments since 1990 (VRBA et al., 2003). Despite similar climatic and morphological conditions, there are significant differences in the N and P fluxes within the ecosystems, as well as in rates of their biological recovery (KOPÁČEK et al., 2002c; MAJER et al., 2003). The role of the different bedrock and soils on these processes have been recently studied (KAŇA & KOPÁČEK, 2006; KOPÁČEK et al., 2006a, b). However, the important part of the catchment ecosystem, the vegetation layer, was neglected until now, because of the lack of reliable data. This study presents the first analysis of the understory vegetation and related element pools for these ecosystems. Here we evaluate: (1) differences in the species composition and biomass of the understory vegetation and the associated pools of major nutrients (C, N, P, Ca, Mg, and K) and ecologically important metals (Al, Fe, Mn) in the catchments of the PL and CT lakes, (2) spatial variability of the studied characteristics (biomass and element pools) within the catchments, and (3) a comparison of the biomass and element pools in the catchments to other available data from Central European spruce forest ecosystems. A comparison of element pools associated with the understory vegetation to those in the soil and tree layers are given in SVOBODA et al. (2006). The role of the understory vegetation in the cycling of terrestrial elements is evaluated by ŠANTRŮČKOVÁ et al. (2006).

Study sites

The research was carried out in the catchments of Plešné Lake (PL; 48°46'35" N, 13°52'0" E; elevation of 1087–1378 m a.s.l.; total forested area of 59.5 ha) and Čertovo Lake (CT; 49°9'55" N, 13°11'50" E; elevation of 1027–1343 m a.s.l.; total forested area of 81.2 ha) in the Bohemian Forest (Šumava, Böhmerwald). The PL catchment is nearly completely covered with forest (~90% of the catchment). The forest stand is on average 160 years old and dominated (99%) by Norway spruce (*Picea abies*), with minor contributions of mountain-ash (*Sorbus aucuparia*) and beech (*Fagus sylvatica*). Timber biomass ranges between 15 and 720 m³ ha⁻¹ (catchment average of 230 m³ ha⁻¹) according to the Stožec, 1995–2004, and Plešný, 1996–2005 forest management plans (I. VICENA – pers. commun.). Only few data are available in the literature on the history of land-use in the PL catchment, summarised by VESELY (1994).

Disturbances occasionally affecting the surroundings of the PL catchment (logging, pasturing, or fires) were probably negligible within the catchment during the last ~250 years (I. VICENA – pers. commun.). Details on forest development in the study area are given by JANKOVSKÁ (2006). The bedrock of the PL catchment is granite, soils are developed from till, rich in sand (~75%), and consist of ~0.2 m deep leptosol (38%) and ~0.45 m deep podsol (29%) or dystric cambisol (27%). The rest of the watershed is rocky; wetlands are negligible (~1%). Soil pH (CaCl₂ extractable) is low, with minimum values of 2.5–3.1 in A-horizons and maximum (3.2–4.4) in low mineral horizons. The mean effective cation exchange capacity of the soils is dominated by exchangeable Al (57%) and protons (28%), while base saturation is 15% (KOPÁČEK et al., 2002a).

The Čertovo Lake catchment is covered with 90–150 year-old Norway spruce forest (*Picea abies*) of at least secondary origin, with sparse European beech (*Fagus sylvatica*). Timber biomass is (according to the Železná Ruda 2003 forest management plans; I. VICENA – pers. commun.) on average 287 m³ ha⁻¹. The land use history of the CT catchment is summarised by VESELY (1994), and suggests important timber harvesting and charcoal and potash production from the Middle Ages to the late 19th century. The bedrock of the CT catchment is made up of mica-schist (muscovitic gneiss), quartzite, and small amounts of pegmatite (VESELY, 1994). The watershed is covered with ~0.5 m deep dystric cambisol (58%), podsol (21%), and shallow (~0.2 m) leptosol (17%); wetlands and bare rocks represent ~3% and 1%, respectively. Fine soil is sandy (48–81%) with a low (1–4%) content of clay. Soil pH (CaCl₂ extractable) is low, with minimum values of 2.5–3.3 in A-horizons and maximum values of 3.6–4.5 in mineral horizons. The mean effective cation exchange capacity of the soils is dominated by exchangeable Al (62%) and protons (29%), while base saturation is 9% (KOPÁČEK et al., 2002b).

As determined from typological maps (using the Czech typological system; VIEWEGH et al., 2003), the CT catchment belongs to the spruce–beech vegetation zone (81% of forested area) and spruce vegetation zone (19% of forested area). The PL catchment belongs to the spruce–beech vegetation zone (12%) and spruce vegetation zone (88%). The distribution and fractions of the groups of the forest types presented within the catchment are shown in Figs 1a and 1b.

Material and methods

Sampling and analysis of the understory vegetation

The investigation of understory vegetation was carried out using the following steps: (1) Plant communities (using plant coenological relevés) were inspected and the boundaries between the distinguished plant communities surveyed. (2) All important types of plant microcoenoses within the plant communities (sensu MATEJKA, 1992b), based on the dominant plant species, were determined by field reconnaissance. (3) Each type of microcoenose within each plant community was sampled to determine aboveground (AG) and belowground (BG) biomass. (4) The area of each plant community was calculated (total area of plant communities gave the total catchment area). (5) Total biomass and catchment weighted mean (CWM) biomass were calculated. (6) Selected elements were analyzed in dried biomass samples (or mixed samples). (7) Total pools of these elements and CWM

pools were calculated using total species biomass and mean element concentration in above-ground and below-ground biomass.

Biomass of the understory vegetation was estimated in both catchments using a combination of the phytosociological survey and biomass sampling during July and August 2004. The forest site typology maps, available at the Regional Plan of Forest Development (prepared by The Forest Management Institute, Brandýs n. Labem), were used as the basis for further plant coenological surveys. According to these maps, the main sets of forest types were distinguished and their area analyzed in GIS. The sets of forest types characterize specific site conditions and therefore a certain composition of plant species (VIEWEGH et al., 2003). After that, a basic field survey analyzing the dominant microcoenoses and plant communities, and the variability in plant species composition within the plant communities, was carried out. In the next step, 35 and 23 plant coenological relevés distributed within the dominant plant communities were recorded in the CT and PL catchments, respectively. The phytosociological surveys were carried out using standard methods (e.g., MORAVEC et al., 1994). Plant nomenclature follows KUBÁT et al. (2002). Each relevé was a circle plot with area of about 200 m². The relevés were placed randomly within the catchments and plant communities. The number of relevés placed in the individual plant communities was proportional to their area. Any distinct change in cover and composition of the plant species encountered during both the basic and phytosociological surveys was marked on the map.

Abundance of the individual plant species was used to distinguish between the plant species that were analyzed for element concentrations. Plant species were not analyzed if their mean abundance was less than 1% in the whole catchment. The following plant species had a mean abundance > 1% in both catchments: *Athyrium distentifolium*, *Avenella flexuosa*, *Calamagrostis villosa*, *Luzula sylvatica*, and *Vaccinium myrtillus*. These species created the main microcoenoses and plant communities, and the understory vegetation in both catchments was finely-structured according to these dominant species.

A 0.5 × 0.5 m frame was used to sample (AG) biomass of the understory vegetation. The frames were randomly placed within both catchments to cover variability in site and stand conditions. The number of biomass samples for individual plant species depended on their relative abundance in each of the catchments. Biomass samples were assigned to microcoenose types and plant communities according to the dominant plant species. For each sample site, AG plant material was cut using a pair of scissors, put into a plastic bag and labelled. The AG biomass samples were brought to the laboratory and individual plant species separated. The AG biomass of *V. myrtillus* was separated into leaves, annual shoots, and woody stems. In this way, the annually produced biomass of *V. myrtillus* was estimated, and the production of AG biomass between the perennial plant species (*V. myrtillus*) and annual plant species (the other species in the catchments) were compared. The samples were dried at 105°C till their weight was stabilized.

A steel corer with a 6.5 cm diameter was used to sample BG biomass. Depth of the soil cores depended on local soil properties (e.g., content of bigger stones). The overall mean depth of the soil cores was about 15 cm; i.e. soil cores contained the majority of the fine root biomass. After

sampling, the material was withdrawn from the steel corer and put into a plastic bag. The BG biomass samples were cleaned in the laboratory using running water on a set of sieves > 0.25 mm in diameter. They were then divided into the following categories: non-woody fine roots, woody roots, and rhizomes. After that, the samples were dried at 105°C until their weight was stabilized.

The following sampling design was used for *Athyrium distentifolium*, which grew in large tussocks. In the area with a high abundance of this species, two rectangular plots (12 and 16 m²) were set only in the CT catchment. All tussocks of *A. distentifolium* in the grid plots were counted and their diameter measured. Then, the AG biomass of several tussocks over a range of diameters was sampled in both catchments. The BG biomass of *A. distentifolium* was sampled in the centre, middle and edge of average-sized tussocks.

All mass and chemical results further reported in this paper are given on a dry weight biomass basis.

Chemical analysis of the biomass

The dry biomass was analysed for total content of the following elements. Total P was determined from a HNO₃ and HClO₄ acid digest using the phosphomolybdate blue method (KOPÁČEK et al., 2001). Carbon (C) and nitrogen (N) were determined by a CN analyser (NC 2100, ThermoQuest, Italy). The total concentration of metals (Ca, Mg, Na, K, Al, Fe, and Mn) was analysed using a H₂SO₄, HNO₃, and HF mixed acid digest (200°C, 2 h) by flame atomic absorption spectrometry.

Statistical analyses

The phytosociological relevés were classified using TWINSPAN (HILL, 1979). The types of plant communities were distinguished according to TWINSPAN classification accounting 3rd classification level. The main types of plant communities (vegetation groups) were determined based on the result of this procedure. The results of the phytosociological survey within the types of plant communities, the distribution of the groups of plant communities in the catchments, and the map notes made during the surveys were used in GIS to analyse and distinguish borders and areas of individual plant communities. The total area of the plant communities groups was later used to calculate the total biomass and CWM biomass of the understory vegetation in each catchment.

Dry weight values of AG biomass of analyzed plant species samples, except *A. distentifolium*, were used to calculate the mean AG biomass individual plant species, which was calculated using equation (1) (competition model of co-existence of the selected plant species) (MATEJKA, 1992a):

$$Af \cdot a^{-1} + Cv \cdot c^{-1} + Ls \cdot l^{-1} + Vm \cdot v^{-1} = 1, \quad (1)$$

where *a*, *c*, *l*, *v* are the values of the "carrying capacity" representing the maximum weight of the biomass (g m⁻²) of the species growing in a monocoenose, and *Af* (*A. flexuosa*), *Cv* (*C. villosa*), *Ls* (*L. sylvatica*), *Vm* (*V. myrtillus*) are the mean dry weights of biomass (g m⁻²) of the species in the given sample. The least square method was used to calculate a regression model. The term carrying capacity is used here in a similar meaning as by ODUM (1959), giving the maximum value of species biomass achievable under optimal conditions. In our sense, it means without the presence of any other competing species. AG biomass values of

A. distentifolium were calculated using a regression model, where the weight of AG biomass (m ; g) is expressed as a function of tussock diameter (D ; cm):

$$m = a \times D^b, \quad (2)$$

where a and b represent result of the regression solution.

The carrying capacity values for individual plant species (or mean biomass of *A. distentifolium*), in other words the biomass of i -th species per unit area – B_i , cover of i -th species in the j -th plant community type – C_{ij} , and the area of the j -th plant community type – A_j (the sum of area of all plant communities gives the total catchment area), were used to calculate the CWM of AG biomass of the i -th species in both catchments:

$$CWM_i = B_i \times \sum_j C_{ij} \times A_j / \sum_j A_j \quad (3)$$

This catchment weighting generates a single value for each parameter, representing a hypothetical situation when all representatives of understory vegetation are uniformly distributed over the whole catchment. The total CWM AG biomass was calculated over all incorporated species.

The mean BG biomass was calculated for all types of microcoenoses. The final BG biomass means have therefore limited significance. The CWM of BG biomass was calculated as a function of mean BG biomass in the i -th type of microcoenose – B_i , relative representation of the i -th type of microcoenose in the j -th plant community type – R_{ij} , and the area of the j -th community type – A_j (whole catchment area was divided into areas of plant community types):

$$CWM = \sum_{i,j} B_i \times R_{ij} \times A_i / \sum_j A_j \quad (4)$$

Total element pools were calculated as a function of the CWM and mean element concentrations of AG and BG biomass (according to individual plant species) for each of the catchments.

Results

Plant coenological surveys

Clear differences were found between the plant communities in the catchments of both lakes. *C. villosa* was the most abundant species in the CT catchment, while *V. myrtillus* was the most abundant species in the PL catchment (Tab. 1). There were 5 main types of understory vegetation plant communities in each of the catchments. Groups *000, *001, and *111 were dominated by *A. distentifolium* and represented the association *Athyrio alpestris-Piceetum* Hartman ex Hartmann et Jahn, 1967 on wet sites. Other groups of plant communities with various shares of *C. villosa* and *V. myrtillus* belonged to the broad association *Calamagrostio villosae-Piceetum* Hartman et Jahn, 1967 (compare HUSOVÁ et al., 2002). Types *010 and *011 surrounding CT Lake were classified as the association *Calamagrostio villosae-Fagetum* Mikyška,

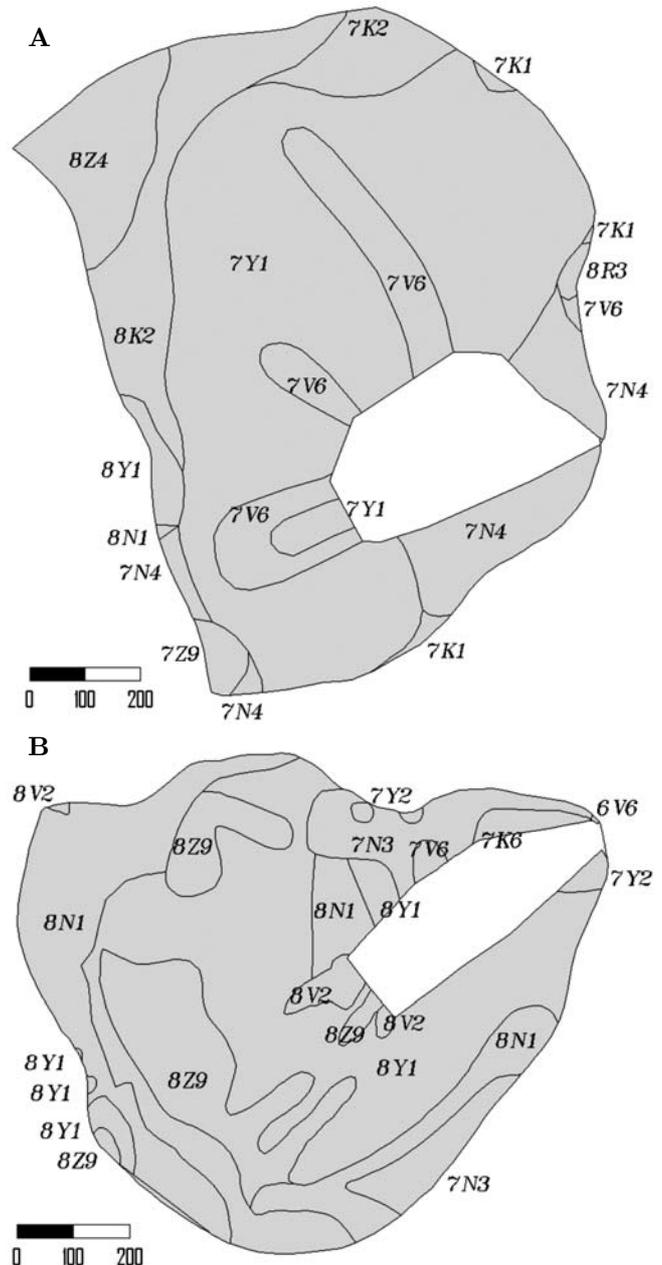


Fig. 1. A – Map of forest types in the catchments of Čertovo Lake. The share of the individual forest type group in the CT catchment was: 7Y – *Fageto-Piceetum saxatilis* – 57.4%, 7K – *Fageto-Piceetum acidophilum* – 4.4%, 7V – *Fageto-Piceetum acerosum humidum* – 9.7%, 8Z – *Sorbeto-Piceetum (humilis)* – 6.8%, 7N – *Fageto-Piceetum lapidosum acidophilum* – 8.9%, 8N – *Piceetum lapidosum acidophilum* – 0.1%, 8Y – *Piceetum saxatile* – 1.3%, 7Z – *Fageto-Piceetum humilis* – 1.0%, 8R – *Piceetum turfosum (montanum)* – 0.4%, 8K – *Piceetum acidophilum* – 10.1%. The terminology of the forest type groups is according to VIEWEGH et al. (2003).

B – Map of forest types in the catchments of Plešné Lake. The share of the individual forest type group in the PL catchment was: 7K – *Fageto-Piceetum acidophilum* – 1.2%, 7N – *Fageto-Piceetum lapidosum acidophilum* – 9.3%, 7V – *Fageto-Piceetum acerosum humidum* – 0.5%, 7Y – *Fageto-Piceetum saxatilis* – 0.9%, 8N – *Piceetum lapidosum acidophilum* – 25.3%, 8V – *Acereto-Piceetum humidum* – 1.6%, 8Y – *Acereto-Piceetum humidum* – 45.0%, 8Z – *Sorbeto-Piceetum (humilis)* – 16.2%. The terminology of the forest type groups is according to VIEWEGH et al. (2003).

Table 1. Mean abundance of plant species of the understory vegetation according to plant community classification groups, as determined by TWINSpan. Species coverage is given in percents.

Catchment	CT	CT	CT	CT	CT	PL	PL	PL	PL	PL
TWINSpan plant community classification groups	*000	*010	*011	*100	*101	*001	*100	*101	*110	*111
Number of relevés	3	14	7	4	7	6	2	12	1	2
Representative area (ha)	7.7	45.8	12.0	2.7	13.0	17.9	1.2	29.3	8.8	2.4
<i>Athyrium distentifolium</i>	32		0.1			27	2.5	0.8	10	35
<i>Avenella flexuosa</i>		16	6.5	16	3.4	4.2	20	1		
<i>Blechnum spicant</i>	0.5	0.5	1		0.2					
<i>Calamagrostis villosa</i>	38	57	37	15	1.2	13	7.5	0.8	1	28
<i>Dryopteris dilatata</i>	5	3.1	2.2	1.5	1.1	10	1.5	1.7	2	
<i>Epilobium angustifolium</i>										1.5
<i>Equisetum sylvaticum</i>	0.3									
<i>Homogyne alpina</i>	0.7	0.1	0.1	0.1		0.4		0.1		
<i>Lastrea limbosperma</i>	10	0.4	0.1							
<i>Luzula luzuloides</i>						0.1				
<i>Luzula sylvatica</i>	6	2.8	1.7	0.1	0.1	11	0.5	0.3		
<i>Lycopodium annotinum</i>	0.3	0.5	0.1		0.4					
<i>Lycopodium clavatum</i>									0.1	
<i>Maianthemum bifolium</i>		0.1	0.3		0.2	0.3		0.1		
<i>Melampyrum sylvaticum</i>		0.1	0.1	0.1	0.1					
<i>Mycelis muralis</i>									0.1	
<i>Oxalis acetosella</i>	1.7	0.6	0.5	0.3		3.7	2	1.5		
<i>Phegopteris connectilis</i>	0.3		0.1			0.7				
<i>Phegopteris dryopteris</i>	0.5									
<i>Polygonatum multiflorum</i>						0.1				
<i>Prenanthes purpurea</i>	0.5	0.1	0.6		0.2	0.2				
<i>Rubus</i> spp. Div.	0.3		0.1			0.5		0.1	0.1	1
<i>Senecio nemorensis</i>				0.1						
<i>Silene vulgaris</i>		0.1								
<i>Soldanella montana</i>						0.2				
<i>Trientalis europaea</i>	0.2	0.1	0.2			0.1	0.5			
<i>Vaccinium myrtillus</i>	1.3	1.1	27	58	75	17	52	82	72	20
<i>Vaccinium vitis-idaea</i>				0.1	0.4	0.2	2.5	2.6	5	2

Explanations: CT – Čertovo Lake; PL – Plešné Lake; *000, *111, and *001 – plant communities dominated by *A. distentifolium*; *010, *011, and *111 – plant communities dominated by *C. villosa*; *100, *101, and *110 – plant communities dominated by *V. myrtillus*).

1972 (MORAVEC et al., 1982; MORAVEC et al., 2000). The spatial distribution of individual vegetation groups within the CT and PL catchments (Fig. 2) was comparable with the forest types distribution (Fig. 1).

Biomass of the understory vegetation

A total of 40 and 50 biomass samples were analysed in the CT and PL catchments, respectively. The biomass of individual samples was highly variable. The mean biomass of *A. flexuosa* was 88 and 160 g m⁻² in the CT and PL catchments, respectively, while it was 154 and 198 g m⁻² for *C. villosa* (Tab. 2). The mean biomass of *L. sylvatica* was 278 and 226 g m⁻² in the CT and PL catchments, respectively, while it was 561 and 713 g m⁻² for *V. myrtillus* (Tab. 2). The estimated biomass of an average-sized *A. distentifolium* tussock (1.4 m diameter) ranged from 187 to 205 g in the CT and PL catchments, respectively. Estimated AG biomass of *A. distentifolium* ranged from 191 to 234 g m⁻². Mean AG biomass was estimated as 213 g m⁻² in both catchments. The mean biomass of *C. villosa* and *L. sylvatica* was similar in the catchments, while those of *A. distentifolium* and *V. myrtillus* were relatively different.

Biomass allocation to the aboveground plant parts of *V. myrtillus* was as follows: leaves – 15% and 11%, annual shoots – 38% and 47%, and woody stems – 47% and 42% in the CT and PL catchments, respectively (Tab. 2).

The BG biomass of woody roots and rhizomes varied greatly among individual species and between the catchments (Tab. 3). Mean fine root biomass also differed among species and between catchments, but was not as variable as for woody roots and rhizomes (Tab. 3). It is likely that the BG fine root biomass values for individual species are within the same range in the CT and PL catchments.

The CWM AG biomass of the understory vegetation recorded in this study was 288 and 730 g m⁻² in the CT and PL catchments, respectively (Tab. 4). The difference in the CWM AG biomass between the catchments was caused by the much higher abundance of *V. myrtillus* in the PL catchment. About 680 g m⁻² of the CWM AG biomass in the PL catchment (730 g m⁻²) was represented by *V. myrtillus*. The woody stems of this evergreen dwarf shrub accounted for 41–47% of AG biomass in the PL catchment (Tab. 2). However,

Table 2. Aboveground biomass according to the main types of microcoenoses.

Locality	Type of microcoenose	N	Mean biomass (g m ⁻²)	Carrying capacity for dominant species (g m ⁻²)	Leaves (%)	Annual shoots (%)	Woody stems (%)
CT	<i>Avenella flexuosa</i>	5	88	95			
CT	<i>Calamagrostis villosa</i>	23	154	219			
CT	<i>Luzula sylvatica</i>	4	278	282			
CT	<i>Vaccinium myrtillus</i>	12	561	924	15.0	37.7	47.3
PL	<i>Avenella flexuosa</i>	4	160	178			
PL	<i>Calamagrostis villosa</i>	6	198	222			
PL	<i>Luzula sylvatica</i>	5	226	231			
PL	<i>Vaccinium myrtillus</i>	20	713	1178	11.0	47.2	41.7

Explanations: CT – Čertovo Lake; PL – Plešné Lake; N – number of samples.

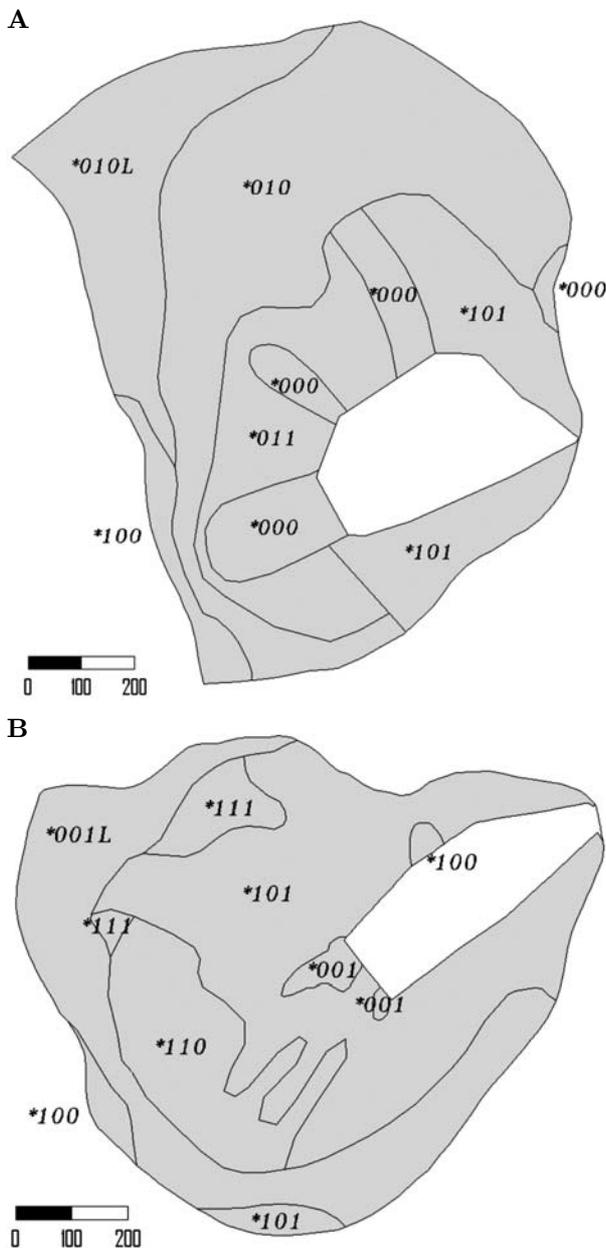


Fig. 2. Map of the plant communities according to TWINSpan classification (Tab. 1) in the catchments of Čertovo Lake (A) and Plešné Lake (B). For explanation of codes of plant communities see Table 1.

the annually produced AG biomass was comparable for both catchments (142 and 124 g m⁻² in the CT and PL catchments, respectively). The CWM BG fine root biomass recorded in this study was 491 and 483 g m⁻² in the CT and PL catchments, respectively (Tab. 5). The CWM biomass of woody roots and rhizomes were not determined, because of high variability in the original data.

Chemical elements in the biomass

Mean element concentrations differed between the AG and BG biomass of the understory vegetation in both catchments (Tab. 6). There were differences also within and between the individual plant species in both catchments. However, the original data were highly variable, with only a relatively low number of samples. The CWM element pools in the AG biomass were 2–3-fold higher in the PL than for the CT catchment, with the greatest differences observed for Ca, Na, and Al (Tab. 7). This difference reflected the higher CWM AG biomass of the understory vegetation in the PL catchment. The evergreen tissues of the dominant *V. myrtillus* in the PL catchment contained higher amounts of elements compared to the biomass of annual plant species that are dominant in the CT catchment. However, the recorded values were similar in both catchments when only element pools stored in annually produced AG biomass were compared (Tab. 7). The CWM element pools stored in the BG biomass of the CT and PL catchments were similar for most of the elements, except Al and Fe. The higher CWM Al and Fe pools in the CT catchment are probably related to the greater abundance of *C. villosa* and its higher concentrations of Al and Fe.

Discussion

Plant communities

There were several possible explanations for the difference in the abundance of the dominant plant species of understory vegetation in the two catchments. The catchments differed in terms of species composition of the natural forest vegetation and soil types. This

Table 3. Below-ground biomass (fine roots, woody roots and rhizomes) in selected sample plots representing the main types of microcoenoses. All values are in g m⁻².

Catchment	Type of microcoenose	N	Fine roots	Woody roots	Rhizomes
CT	<i>Athyrium distentifolium</i>	6	669	28	nd
CT	<i>Avenella flexuosa</i>	5	785	356	8
CT	<i>Calamagrostis villosa</i>	23	567	59	190
CT	<i>Luzula sylvatica</i>	4	321	666	137
CT	<i>Vaccinium myrtillus</i>	12	730	212	1
PL	<i>Athyrium distentifolium</i>	6	693	207	924
PL	<i>Avenella flexuosa</i>	4	349	125	3
PL	<i>Calamagrostis villosa</i>	6	391	481	113
PL	<i>Luzula sylvatica</i>	5	376	199	18
PL	<i>Vaccinium myrtillus</i>	20	616	461	95

Explanations: CT – Čertovo Lake; PL – Plešné Lake; N – number of samples.

Table 4. Above-ground biomass of the understory vegetation according to plant communities for species *A. flexuosa* (Af), *C. villosa* (Cv), *L. sylvatica* (Ls), *V. myrtillus* (Vm) and *A. distentifolium* (Ad).

A. Catchment of Čertovo Lake.												
Plant community	Dominant species carrying capacity (g m ⁻²)	Af	Cv	Ls	Vm	Ad	Af	Cv	Ls	Vm	Ad	Total
	Total area (m ²)	Species representation (%)					Average biomass (g m ⁻²)					
		94.5	219.5	282.1	923.7	213						
*000	77000		38	6	1	35		83	16.9	9.2	74.6	
*010	320570	16	57		1		15.1	125		9.2		
*010L	137460	16	57	8	1		15.1	125	22.6	9.2		
*011	120400	7	37	2	27		6.6	81	5.6	249		
*100	26550	16	15		58		15.1	33		536		
*101	129980	4	1		75		3.8	2.2		693		
CWM AG biomass (g m ⁻²)							10.6	92	6.3	172	7.1	287
B. Catchment of Plešné Lake.												
Plant community	Dominant species carrying capacity (g m ⁻²)	Af	Cv	Ls	Vm	Ad	Af	Cv	Ls	Vm	Ad	Total
	Total area (m ²)	Species representation (%)					Average biomass (g m ⁻²)					
		178.3	222.4	230.9	1177.0	193						
*001	8940	4	13		17	27	7.1	28.9		200	52.1	
*001L	170370	4	13	15	17	27	7.1	28.9	34.6	200	52.1	
*100	11890	20	8		52	2	35.7	17.8		612	3.9	
*101	292640	1	1		82	1	1.8	2.2		965	1.9	
*110	87670		1		72	10		2.2		847	19.3	
*111	24480		28		20	35		62.3		235	67.6	
CWM AG biomass (g m ⁻²)							3.7	13	9.9	681	22.3	730

was reflected by the phytosociological classification, where subassociation *Calamagrostio villosae-Piceetum typicum* Hartmann in Hartmann et Jahn, 1967 was dominant in the CT catchment (with dominant *C. villosa*), while subassociation *Calamagrostio villosae-Piceetum vaccinietosum myrtilli* Jirásek, 1996 was dominant in the PL catchment (with dominant *V. myrtillus*). The soils of the CT catchment were mostly cambisols, while the PL catchment was dominated by undeveloped organic rich soils (KOPÁČEK, 2002a, b). The latter soil type is often dominated by *V. myrtillus*.

The catchments also had different disturbance histories. Based on the phytosociological classification, a significant part of the forest stands of the CT catchment would be dominated by beech in the absence of human impact. However, the forest stands in this catchment were logged in the past and the species composition of the tree layer was probably changed, being now dominated by spruce (VESELÝ, 1994). The forest stands of the PL catchment are more or less naturally dominated by spruce. There is evidence that the coverage of grasses could increase, while that of dwarf shrubs would decrease, following man-made disturbance (UOTILA &

Table 5. Below-ground biomass of fine roots of the understory vegetation according to plant communities for the microcoenoses dominated by *A. flexuosa* (Af), *C. villosa* (Cv), *L. sylvatica* (Ls), *V. myrtillus* (Vm) and *A. distentifolium* (Ad) in the catchment of Čertovo Lake.

A. Catchment of Čertovo Lake.												
	Dominant species carrying capacity (g m ⁻²)	Af	Cv	Ls	Vm	Ad	Af	Cv	Ls	Vm	Ad	Total
Plant community	Total area (m ²)	Species representation (%)					Average biomass (g m ⁻²)					
		785.2	567.3	320.9	730	668.5						
*000	77000		38	6	1	35		216	19.2	7.3	234	
*010	320570	16	57		1		126	323		7.3		
*010L	137460	16	57	8	1		126	323	25.7	7.3		
*011	120400	7	37	2	27		55	210	6.4	197		
*100	26550	16	15		58		126	85		423		
*101	129980	4	1		75		31.4	5.7		548		
CWM BG biomass (g m ⁻²)							88.2	238	7.1	136	22.2	491
B. Catchment of Plešné Lake.												
	Dominant species carrying capacity (g m ⁻²)	Af	Cv	Ls	Vm	Ad	Af	Cv	Ls	Vm	Ad	Total
Plant community	Total area (m ²)	Species representation (%)					Average biomass (g m ⁻²)					
		348.7	390.9	375.6	616.4	693.3						
*001	8940	4	13		17	27	14	50.8		105	187	
*001L	170370	4	13	15	17	27	14	50.8	56.3	105	187	
*100	11890	20	8		52	2	69.7	31.3		321	13.9	
*101	292640	1	1		82	1	3.5	3.9		506	6.9	
*110	87670		1		72	10		3.9		444	69.3	
*111	24480		28		20	35		109		123	243	
CWM BG biomass (g m ⁻²)							7.3	22.9	16.1	357	80.2	483

KOUKI, 2005). The dense canopy of the beech stands results in often sparse understory vegetation, while the canopy of the spruce stands lets through more light, which enables the development of light demanding grass species such as *C. villosa* (HOLEKSA, 2003; WILD et al., 2004).

Acid deposition, resulting in soil acidification in the catchments of both lakes, was another important factor that could possibly affect species composition (KOPÁČEK, 2002c). There is some evidence that long-term soil acidification may cause changes in forest understory vegetation (KUBÍČEK et al., 1989; TONJE et al., 2004). Nitrogen deposition was shown to be the main factor affecting the vitality of dwarf shrubs not only in boreal forests of Fennoscandia (NORDIN et al., 1998; STRENGBOM et al., 2002, 2003, 2004). Their decline was followed by an increase in grass coverage induced by increased light availability. The fate of N differed between the PL and CT catchments, with the CT catchment being more saturated by N than the PL catchment. But, due to the dominant effect of light availability over N deposition on the composition of understory vegetation (STRENGBOM et al., 2004), the historical changes in forest structure and composition had probably a greater effect on the development of understory vegetation than N deposition in the catchments studied.

Biomass

The mean biomass of individual understory species is within the range found in the available literature. The mean AG biomass of *C. villosa* found in this study was 154 and 198 g m⁻² in the CT and PL catchments, respectively. The values of the mean AG biomass for this species in Central Europe vary from 70 to 323 g m⁻² (FIEDLER & HOHNE, 1987; MATĚJKA, 1992a; PYŠEK, 1993; JAKRLOVÁ, 1996) in forest stands with closed canopy. In contrast, higher biomass was reported for this species (321–726 g m⁻²) from areas with open canopy or on deforested sites (FIALA et al., 1989; JAKRLOVÁ, 1996). The mean BG biomass of *C. villosa* found in this study was 567 and 391 g m⁻² in the CT and PL catchments, respectively. The mean BG biomass of this species reported by other studies was highly variable, from 48 to 3461 g m⁻² (FIEDLER & HOHNE, 1987; FIALA et al., 1989; PYŠEK, 1993). The differences depended on whether the sampling was done in open or closed canopy forests, and the reported part of BG biomass (fine roots or rhizomes). The mean AG biomass of *V. myrtillus* found in this study (561 and 713 g m⁻² in the CT and PL catchments, respectively) is at the low end of reported values for this species (100 and 3700 g m⁻²) (GERDOL et al., 2004). The relatively high mean biomass of *V. myrtillus* in the PL catchment is probably related to the more open canopy of

Table 6a. Mean element concentrations in the biomass.

Catchment	Species/category	Type of microcoenose	N	Mean/ SD	C mol	N kg ⁻¹	P	Ca	Mg	mmol kg ⁻¹				
										Na	K	Al	Fe	Mn
CT	Aboveground biomass													
	<i>Athyrium distentifolium</i>	Ad	4	Mean SD	40 0.3	2.1 0.2	81 24	83 13	107 5.3	0.5 0.2	920 72	3.2 0.4	1.6 0.1	9.4 2.9
	<i>Avenella flexuosa</i>	Af	4	Mean SD	40 0.3	1.7 0.1	62 2	24 2.7	48 1.5	1 0.2	396 44	4.9 2	2.4 0.3	4 0.9
	<i>Calamagrostis villosa</i>	Ls	1		41	1.8	78	25	25	0.5	461	1.1	1.2	6.2
		Cv	7	Mean SD	40 0.7	1.6 0.1	60 6.8	27 3.7	36 4.7	0.5 0.2	429 44	2.2 0.4	1.7 0.1	5.4 1.5
	<i>Luzula sylvatica</i>	Ls	4	Mean SD	39 0.3	1.5 0.1	84 11	28 1.6	77 11	1.5 0.6	559 49	5.1 1.5	2 0.2	6.4 1
	<i>Vaccinium myrtillus</i> – L	Vm	4	Mean SD	42 0.9	1.5 0.1	49 13	157 14	81 15	0.6 0.0	234 63	6.9 0.8	1.8 0.2	14 4.7
	<i>Vaccinium myrtillus</i> – A	Vm	4	Mean SD	43 0.4	0.8 0.1	27 5.6	127 16	35 4.3	1 0.2	88 17	5.9 0.3	1.6 0.1	18 7.6
	<i>Vaccinium myrtillus</i> – W	Vm	4	Mean SD	42 0.5	0.5 0.1	14 3.5	55 13	14 3	0.4 0.1	38 7.3	6.3 0.6	1.7 0.1	11 4.1
	CT	Below-ground biomass												
Fine roots		Af	3	Mean SD	43 0.5	1 0.1	26 0.6	69 15	29 3.1	8.2 1.6	34 9.6	72 11	21 3.5	1.2 0.1
		Cv	9	Mean SD	43 1.3	1 0.1	28 5	67 23	27 5.7	8 3.5	13 31	122 87	32 19	1.8 0.6
		Ls	2	Mean SD	42 0.3	1 0.0	31 1.2	81 6.2	33 0.4	9.6 2.3	27 6.2	102 21	41 5	1.9 0.2
		Vm	4	Mean SD	44 0.7	0.8 0.1	22 3	60 13	23 5.7	4.8 2	26 7.1	62 24	18 5.8	3.9 2.8
Fine roots – between tussocks		Ad	1		43	1.2	29	81	58	7	24	166	34	1.8
Rhizomes		Cv	4	Mean SD	40 0.6	0.9 0.1	30 11	27 4	12.2 2	5.5 1.5	66 27	35 15	8.3 2.5	1.5 0.5

Explanations: N – number of samples; Mean – arithmetic mean; SD – standard deviation. Types of microcoenoses according to dominant species: Af – *A. flexuosa*; Cv – *C. villosa*; Ls – *L. sylvatica*; Vm – *V. myrtillus*; Ad – *A. distentifolium*. Aboveground categories for *V. myrtillus*: L – leaves; A – annual shoots; W – woody shoots. CT – Čertovo Lake.

the forest stand; more light was available in the upper part of the catchment. Site conditions (soil type and tree canopy) were the main factors that influenced the biomass of this species. The mean AG biomass of *A. flexuosa* found in this study was 88 and 161 g m⁻² in the CT and PL catchments, respectively, and was similar to values reported by MATĚJKA (1992a) (134–151 g m⁻²) and PALVIAINEN et al. (2005a) (40–90 g m⁻²). There is no simple explanation for the difference in mean biomass of *A. flexuosa* between the CT and PL catchments, because of the relatively low number of samples and their high variability. The recorded values of AG biomass (278 and 226 g m⁻²) of *L. sylvatica* were similar to ones reported by MATĚJKA (1992a) (192 g m⁻²). In spite of these data, there are not many studies dealing with the biomass of different understory vegetation species. However, it is still possible to conclude that forest canopy status (open or closed) and site conditions (soil type and fertility) are the main factors that can affect the biomass of the understory vegetation.

The CWM AG biomass for the herb species recorded in this study was 288 and 730 g m⁻² in the CT and PL catchments, respectively. There are only

a few studies that measured the biomass of the understory vegetation on the stand or catchment scales under similar site conditions. The recorded means for AG and BG biomass were 1290 and 516 g m⁻², respectively in a spruce forest in Germany (SCARASCIA-MUGNOZZA et al., 2000). Even though the understory vegetation at this site was composed of similar species as in our sites (*C. villosa*, *A. flexuosa*, and *V. myrtillus*), the total biomass of the understory vegetation was higher. Much lower biomass values were reported from another spruce forest in Germany, where the recorded mean AG and BG values were 107 and 77–188 g m⁻², respectively (GERSTBERGER et al., 2004). The understory vegetation at this German site was composed mostly of *C. villosa*. Similar AG and BG biomass values were reported from an old-growth spruce boreal forest in Finland. The understory vegetation at this site was composed mostly of *V. myrtillus*, which accounted for 150 and 300 g m⁻² of AG and BG biomass, respectively (PALVIAINEN et al., 2005a). The characteristics of the tree stand, soil fertility, soil properties and climate have to be always considered when comparing the biomass of understory vegetation between sites. Understory vege-

Table 6b. Mean element concentrations in the biomass.

Catchment	Species/category	Type of microcoenose		Mean/ SD	C mol kg ⁻¹	N	P	Ca	Mg	Na	K	Al	Fe	Mn	
			N												
PL	Aboveground biomass														
	<i>Athyrium distentifolium</i>	Ad	4	Mean SD	41 0.5	2.1 0.1	136 20	71 23	75 6	0.7 0.5	817 176	4.1 2.5	1.8 0.5	6.1 2	
	<i>Avenella flexuosa</i>	Af	4	Mean SD	40 0.2	1.4 0.1	59 5.9	26 3	45 6.7	0.9 0.0	352 70	4.2 0.4	2 0.1	5.1 1	
	<i>Calamagrostis villosa</i>	Cv	3	Mean SD	39 0.2	1.6 0.0	74.2 7	33 9	37 1.6	0.4 0.2	449 85	1.9 0.2	1.8 0.1	7.1 2.4	
		Ls	1		39	1.4	63.2	18	18	1.2	398	1.1	1.2	1.9	
	<i>Luzula sylvatica</i>	Cv	1		40	1.4	60.1	75	87	1.2	974	6	1.5	5.8	
		Ls	3	Mean SD	40 0.1	1.4 0.1	79.1 15.6	54 8.8	68 5.9	2.2 0.2	546 92	12 2.6	3.4 0.5	6 1.5	
	<i>Vaccinium myrtillus</i> – L	Vm	8	Mean SD	42 0.5	1.5 0.1	54.6 7.8	176 16	84 11	0.5 0.2	255 64	6.5 0.8	1.6 0.2	9.8 2.7	
	<i>Vaccinium myrtillus</i> – A	Vm	8	Mean SD	43 0.5	0.8 0.1	29.2 5.9	137 35	40 9.7	1.2 0.4	102 12	7.1 1.9	1.4 0.2	12 3	
	<i>Vaccinium myrtillus</i> – W	Vm	8	Mean SD	43 0.5	0.5 0.1	15.7 2.8	64 12	18 3.7	0.6 0.1	47 6.7	6.9 1.1	1.7 0.2	9.1 1.9	
	PL	Below-ground biomass													
		fine roots	Af	2	Mean SD	43 0.7	1 0.0	26.2 1.7	56 8.2	19 1.5	14.7 8.6	34 15	83 60	12 3.3	1 0.2
			Cv	2	Mean SD	42 1.0	1 0.0	35.6 7.4	57 12	21.2 3.6	21 2.2	45 3.9	145 37	23 6.5	1.4 0.2
			Ls	2	Mean SD	44 0.3	0.9 0.1	33.2 2.4	72 4.8	21 0.9	11 3	35 12	110 28	20 3.1	2.1 0.3
		Vm	8	Mean SD	42 0.6	0.7 0.2	18.7 3.6	75 24	26 7.7	8 5	23 5.1	43 24	9.1 3.6	3.5 1.7	
Fine roots – in tussock		Ad	1		43	1.3	38.3	86	44	35	34	159	18	1.2	
Fine roots – tussock fringe		Ad	1		42	1.2	34.1	58	26	40	57	304	36	1.4	
Fine roots – between tussocks		Ad	1		44	1.3	38.3	71	50	10	26	135	19	0.8	
Rhizomes – between tussocks		Ad	1		43	1.4	42.2	92	63	7.3	98	19	1.9	0.6	
Rhizomes		C	1		40	0.9	56.3	21	10	22	91	57	8.2	1.5	

Explanations: N – number of samples; Mean – arithmetic mean; SD – standard deviation. Types of microcoenoses according to dominant species: Af – *A. flexuosa*; Cv – *C. villosa*; Ls – *L. sylvatica*; Vm – *V. myrtillus*; Ad – *A. distentifolium*. Aboveground categories for *V. myrtillus*: L – leaves; A – annual shoots; W – woody shoots. PL – Plešné Lake.

tation is heterogeneous and can exhibit wide variability due to numerous factors (JALONEN et al., 1998). The biomass values recorded in this study correspond to the specific site and climate conditions of our study sites. The biomass was higher in the more productive German site, but lower in the less productive Finland site.

Chemical elements in the biomass

Element concentrations in sampled plant tissues differed both within and among species. This is in accordance with the general principles of plant mineral nutrition in that not all plant species require the same complement of minerals in the same amounts, but each species requires minerals in various proportions (CHAPIN & AERTS, 2000). Despite the relatively low number of analysed samples, some differences were found in the element concentrations of plant tissues. For example, there was a clear difference in Al concentrations between the AG and BG biomass of all understory

vegetation species. The increased concentrations of Al in the BG biomass could indicate increased availability of Al in soil solution due to soil acidification (KOPÁČEK, 2002a).

There are a limited number of generally accessible studies dealing with the nutrition of the plant species that we sampled, even though these are common species all over Europe. This relative lack of studies is likely due to the high spatial and seasonal variability in element concentrations in plant tissues (KAUNISTO & SARJALA, 2003; GERDOL et al., 2004). The concentrations of P, Ca, and K were 29, 224, and 196 mmol kg⁻¹, respectively, for *V. myrtillus* growing in a spruce boreal forest in Finland (PALVIAINEN et al., 2005b). The concentrations for *A. flexuosa* at the same site were 62, 65, and 766 mmol kg⁻¹ for P, Ca, and K, respectively. The element concentrations for *V. myrtillus* in our study ranged from 13 to 55 (P), 55 to 176 (Ca), and 38 to 255 (K) mmol kg⁻¹. These concentrations depended on

Table 7. Mean pools of elements calculated per unit area within catchment.

Catchment	Dominant species/ type of microcoenose	Category	Biomass g m ⁻²	C mol m ⁻²	N mol m ⁻²	P	Ca	Mg	Na mmol m ⁻²	K	Al	Fe	Mn
CT	<i>Avenella flexuosa</i>	AG	10.6	0.43	0.02	0.66	0.26	0.51	0.01	4.20	0.05	0.03	0.04
	<i>Calamagrostis villosa</i>	AG	92.0	3.67	0.14	5.56	2.48	3.29	0.04	39.5	0.21	0.16	0.45
	<i>Luzula sylvatica</i>	AG	6.3	0.25	0.01	0.53	0.18	0.49	0.01	3.52	0.03	0.01	0.04
	<i>Vaccinium myrtillus</i>	AG-L	25.7	1.08	0.04	1.27	4.05	2.08	0.02	6.02	0.18	0.05	0.36
	- " -	AG-A	64.7	2.76	0.05	1.72	8.20	2.29	0.06	5.71	0.38	0.09	1.17
	- " -	AG-W	81.1	3.44	0.04	1.12	4.50	1.15	0.03	3.07	0.51	0.14	0.90
	<i>Athyrium distentifolium</i>	AG	7.1	0.28	0.02	0.64	0.59	0.76	0.01	6.53	0.02	0.01	0.07
	Annually produced AG biomass			5.71	0.22	8.65	7.55	7.13	0.08	59.7	0.49	0.25	1.01
	Total AG biomass			11.9	0.31	11.5	20.3	10.6	0.17	68.5	1.38	0.48	3.09
	PL	<i>Avenella flexuosa</i>	AG	3.7	0.15	0.01	0.22	0.09	0.17	0.01	1.30	0.02	0.01
<i>Calamagrostis villosa</i>		AG	13.0	0.50	0.02	0.96	0.43	0.48	0.01	5.84	0.03	0.02	0.09
<i>Luzula sylvatica</i>		AG	9.9	0.39	0.01	0.78	0.54	0.67	0.02	5.41	0.13	0.03	0.06
<i>Vaccinium myrtillus</i>		AG-L	74.9	3.15	0.11	4.09	13.2	6.29	0.04	19.1	0.49	0.12	0.73
- " -		AG-A	321.2	13.7	0.24	9.38	44.0	12.9	0.37	32.8	2.29	0.44	3.82
- " -		AG-W	283.8	12.1	0.15	4.45	18.2	5.11	0.16	13.2	1.96	0.50	2.57
<i>Athyrium distentifolium</i>		AG	22.3	0.91	0.05	3.02	1.58	1.68	0.02	18.2	0.09	0.04	0.14
Annually produced AG biomass				5.11	0.20	9.08	15.8	9.29	0.08	49.8	0.74	0.22	1.04
Total AG biomass				30.9	0.59	22.9	78.0	27.3	0.61	95.9	4.98	1.16	7.43
CT		<i>Avenella flexuosa</i>	fine roots	88.2	3.83	0.09	2.30	6.12	2.59	0.78	2.98	6.32	1.83
	<i>Calamagrostis villosa</i>	fine roots	237.7	10.1	0.23	6.63	15.8	6.32	1.89	7.46	29.1	7.55	0.44
	<i>Luzula sylvatica</i>	fine roots	7.1	0.30	0.01	0.22	0.57	0.24	0.07	0.19	0.72	0.29	0.01
	<i>Vaccinium myrtillus</i>	fine roots	135.5	5.94	0.11	2.92	8.18	3.09	0.66	3.55	8.42	2.40	0.53
	<i>Athyrium distentifolium</i>	fine roots	22.2	0.96	0.03	0.64	1.81	1.28	0.15	0.52	3.69	0.75	0.04
	Total BG biomass			21.14	0.46	12.7	32.5	13.5	3.49	14.7	48.2	12.8	1.13
PL	<i>Avenella flexuosa</i>	fine roots	7.3	0.31	0.01	0.19	0.41	0.14	0.11	0.24	0.61	0.09	0.01
	<i>Calamagrostis villosa</i>	fine roots	22.9	0.97	0.02	0.82	1.30	0.49	0.49	1.02	3.33	0.53	0.03
	<i>Luzula sylvatica</i>	fine roots	16.1	0.71	0.02	0.53	1.16	0.33	0.17	0.56	1.77	0.32	0.03
	<i>Vaccinium myrtillus</i>	fine roots	356.5	15.0	0.26	6.67	26.8	9.07	2.84	8.06	15.3	3.27	1.24
	<i>Athyrium distentifolium</i>	fine roots	80.2	3.44	0.10	2.96	5.74	3.19	2.28	3.11	16.0	1.95	0.09
	Total BG biomass			20.5	0.41	11.2	35.4	13.2	5.89	13.0	36.9	6.15	1.40

Explanations: Category: AG – above-ground biomass (AG-L – leaves, AG-A – annual shoots, AG-W – woody shoots; annual biomass contains total biomass all species except *V. myrtillus* of which only leaves and annual shoots are included). Biomass reports share of plant species on values of AG and BG CWB. CT – Čertovo Lake; PL – Plešné Lake.

the type of biomass sampled (leaves, annual shoots, and woody stems) (Tab. 6). For *A. flexuosa*, the P, Ca, and K concentrations ranged from 59 to 62, 24 to 26, and 352 to 396 mmol kg⁻¹, respectively. N and P concentrations were 0.3 mol kg⁻¹ and 29 mmol kg⁻¹, respectively, in *V. myrtillus* from a sub-alpine forest in Italy (GERDOL et al., 2004). The element concentrations of *V. myrtillus* in our study ranged from 14 to 55 mmol kg⁻¹ and from 0.7 to 1.5 mol kg⁻¹ for P and N, respectively. Given the fact that these three sites differ in their site conditions and the analytical methods used, these values are reasonably comparable.

The CWM element pools differed between the CT and PL catchments. The total amount of elements stored in the PL catchment understory vegetation was considerably higher, because of the higher CWM biomass of its understory vegetation. But the el-

ement pools were similar when only annually produced AG biomass was compared. PALVIAINEN et al. (2005b) showed that total element pools in understory vegetation biomass (504 g m⁻²) was 0.34 mol m⁻² (N), 13 mmol m⁻² (P), 41 mmol m⁻² (K), and 35 mmol m⁻² (Ca) in the boreal spruce forest. The total CWM biomass of the understory vegetation was 288 and 730 g m⁻², with element pools of 0.3 and 0.6 mol m⁻² (N), 12 and 23 mmol m⁻² (P), 69 and 96 mmol m⁻² (K), and 20 and 78 mmol m⁻² (Ca) in the PL and CT catchments, respectively. These reported element pools are within the range found in the boreal forest.

Conclusions

We observed important differences in the character, composition, and biomass of understory vegetation be-

tween the PL and CT catchments in the Bohemian Forest, resulting in significant differences in the associated pools of key elements. The CWM AG biomass of the understory vegetation, as well as the associated CWM elements pools, was 2–3 times higher in the PL than CT catchment, with the greatest differences observed for Ca, Na, and Al. The differences were predominantly caused by higher abundance of *V. myrtillus* in the PL catchment, while the understory vegetation was dominated by *C. villosa* in the CT catchment. The CWM BG biomass of the understory vegetation was similar in both catchments.

The differences in both site factors and history of land use were probably responsible for the observed differences in the character and pools of the understory vegetation in the CT and PL catchments. The catchments differ in the proportion of individual soil types and catchments morphology. Without a more detailed knowledge of the land use history and its impact on the vegetation in the catchments, it is, however, difficult to conclude whether the difference in the character of understory vegetation reflects the natural or anthropogenic factors.

The major aim of this study was to evaluate differences in the understory vegetation in the PL and CT catchments. Further evaluation of the data (SVOBODA et al., 2006; ŠANTRŮČKOVÁ et al., 2006), however, suggested their following implication for whole ecosystems studies: (1) The understory vegetation of *C. villosa* played an important role in N balance on the catchment scale. The higher abundance of *C. villosa* in the CT catchment possibly caused higher N flux from the litter to the CT soil compared to the PL catchment, where *V. myrtillus* was the most abundant species (ŠANTRŮČKOVÁ et al., 2006). (2) Comparing the data on element pools in understory vegetation to those in the tree layer showed that the annual uptake of nutrients by these two vegetation layers were of the same magnitude (SVOBODA et al., 2006). These results imply that adequate attention should be paid to the role of understory vegetation in element cycling at the whole ecosystem scale.

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